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(54) Title: STABLE PROTEIN AND NUCLEIC ACID FORMULATIONS USING NON-AQUEOUS, ANHYDROUS, APROTIC, HYDROPHOBIC, NON-POLAR VEHICLES WITH LOW REACTIVITY		
(57) Abstract <p>This invention relates to stable non-aqueous formulations which are suspensions of proteinaceous substances or nucleic acids in non-aqueous, anhydrous, aprotic, hydrophobic, non-polar vehicles with low reactivity. More specifically, the present invention relates to stable protein or nucleic acid formulations wherein the compound remains in stable, dry powder form, yet the formulation is flowable and, therefore amenable to delivery to an animal via injection, transdermal administration, oral delivery or using an implantable device for sustained delivery. These stable formulations may be stored at elevated temperatures (e.g., 37 °C) for long periods of time and are especially useful as flowable formulations which can be shipped and/or stored at high temperatures or in implantable delivery devices for long term delivery (e.g., 1-12 months or longer) of drug.</p>		

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1 **STABLE PROTEIN AND NUCLEIC ACID FORMULATIONS USING NON-**
2 **AQUEOUS, ANHYDROUS, APROTIC, HYDROPHOBIC, NON-POLAR**
3 **VEHICLES WITH LOW REACTIVITY**

4
5 **FIELD OF THE INVENTION**

6
7 This invention relates to stable non-aqueous formulations of
8 proteins and nucleic acids. The stable formulations of the present
9 invention are suspensions of particles containing proteins or nucleic acids
10 in non-aqueous, anhydrous, aprotic, hydrophobic, non-polar vehicles with
11 low reactivity.

12
13 **BACKGROUND OF THE INVENTION**

14
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16 The following references are referred to by numbers in brackets ([])
17 at the relevant portion of the specification.

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1 The disclosure of each of the above publications, patents or patent
2 applications is hereby incorporated by reference in its entirety to the same
3 extent as if the language of each individual publication, patent and patent
4 application were specifically and individually incorporated by reference.

5

6 Background of the Invention:

7 Peptides, polypeptides, proteins and other proteinaceous
8 substances (e.g., viruses, antibodies), collectively referred to herein as
9 proteins, have great utility as pharmaceuticals in the prevention, treatment
10 and diagnosis of disease. Proteins are naturally active in aqueous
11 environments, thus the preferred formulations of proteins have been in
12 aqueous solutions. However, proteins are only marginally stable in
13 aqueous solutions. Thus, protein pharmaceuticals often require
14 refrigeration or have short shelf-lives under ambient conditions. Further,
15 many proteins have only limited solubility in aqueous solutions. Even
16 when they are soluble at high concentrations, they are prone to
17 aggregation and precipitation.

18 Proteins can degrade via a number of chemical mechanisms,
19 including deamidation of asparagine and glutamine; oxidation of
20 methionine and, to a lesser degree, tryptophan, tyrosine and histidine;
21 hydrolysis of peptide bonds; disulfide interchange; and racemization of
22 chiral amino acid residues [1, 2 and 24-28]. Water is a reactant in nearly
23 all of these degradation pathways. Further, water acts as a plasticizer
24 which facilitates unfolding and irreversible aggregation of proteins. Since
25 water is a participant in almost all protein degradation pathways,
26 reduction of the aqueous protein solution to a dry powder provides an
27 alternative formulation methodology to enhance the stability of protein
28 pharmaceuticals. Proteins can be dried using various techniques,
29 including freeze-drying, spray-drying and dessication. Aqueous solutions
30 of proteins are thus dried and stored as dry powders until their use is
31 required.

1 A serious drawback to drying of proteins is that often one would
2 like to use proteins in some sort of liquid form. Parenteral injection and
3 the use of drug delivery devices for sustained delivery of drug are two
4 examples of applications where one would like to use proteins in a liquid
5 form. For injection, dried proteins must be reconstituted, adding
6 additional steps which are time-consuming and where contamination may
7 occur, and exposing the protein to potentially destabilizing conditions
8 [15].

9 The sustained parenteral delivery of drugs, in particular proteins and
10 nucleic acids, provides many advantages. The use of implantable devices
11 for sustained delivery of a wide variety of drugs or other beneficial agents
12 is well known in the art. Typical devices are described, for example, in
13 U.S. Patents Nos. 5,034,229; 5,057,318; and 5,110,596. The
14 disclosure of each of these patents is incorporated herein by reference.

15 Proteins are only marginally soluble in non-aqueous solvents, and
16 such solvents typically unfold and denature proteins [4, 16].
17 Solubilization of native proteins in non-aqueous solvents typically requires
18 derivatization or complexation of the protein [12]. In attempting to
19 achieve enzymatic catalysis in organic media, Klibanov and others have
20 shown that certain catalytic enzymes can be suspended in non-aqueous
21 vehicles as powders, typically in hydrophilic organic solvents including
22 alcohol ketones and esters [3, 5-11, 13 and 18-23]. With enzyme
23 hydration levels $\geq 10\%$ and/or the addition of low molecular weight protic
24 compounds, these enzymes can have enough conformational mobility to
25 exhibit appreciable enzymatic activity. Optimal activity levels are
26 apparently achieved at enzyme hydration of approximately 30%. At a
27 minimum, such enzymatic activity requires a level of "essential water"
28 hydrating the protein. However, hydration levels (generally 10-40% w/w
29 water/protein) and/or protic solvents, such as those used in these studies,
30 typically result in unacceptable stability of proteins for pharmaceutical
31 purposes. A further requirement for catalysis in non-aqueous solvents is
32 that the enzyme be dried from a solution having a pH near the optimal pH

1 for the enzymatic activity. This pH limitation is detrimental to storage of
2 protein pharmaceuticals, because most protein degradation mechanisms
3 are pH dependent, and it is often the case that proteins are most stable
4 when dried at pH values far from the value where they exhibit bioactivity
5 [1]. Further, such catalytic enzyme systems are not amenable to the
6 addition of protein stabilizers, particularly those that function by hydrogen
7 bonding to the protein and reducing enzyme hydration (e.g.,
8 carbohydrates) [14].

9 The use of perfluorocarbons as components of drug delivery
10 vehicles for certain ophthalmic compositions has been disclosed [29, 30].

11 Similarly, suspensions of growth hormone in triacetin or polyethylene
12 glycol has been published [31].

13 The field of gene therapy or gene transfer is advancing both
14 experimentally and clinically. Nucleic acids have been transferred into
15 cells using viral vectors such as adenovirus, retrovirus, adeno-associated
16 virus, vaccinia virus, and sindbis virus, among others. Non-viral methods
17 have also been used, including calcium phosphate precipitation, DEAE
18 dextran, injection of naked DNA, electroporation, cochleates, cationic lipid
19 complexes, liposomes, polymers (such as dendrimers and PLGA),
20 virosomes, and the like.

21 DNA complexed with cationic lipids and/or liposomes has been
22 shown to be an efficient means of transfecting a variety of mammalian
23 cells. Such complexes are simple to prepare and may be used with a wide
24 variety of DNA's and RNA's with little restriction to the size of nucleic
25 acid. They have the ability to transfect many different cell types with
26 efficiency and are not immunogenic [32, 33, 35, 36]. Current nucleic
27 acid formulations, including DNA/liposome and RNA/liposome complexes,
28 must be mixed shortly before administration, resulting in inconvenience in
29 manufacture, shipping, storage and administration [35, 37-40].

30 Frequently, these two-part formulations are not very highly concentrated,
31 requiring the administration of large volumes of solution. Dry powder
32 formulations containing lyophilized nucleic acid/liposome complexes have

1 also been used [34, 41], but they require reconstitution with suitable
2 aqueous solution just prior to administration. Aqueous complexes are
3 inherently unstable and lose most, if not all, of their transfection activity
4 within hours or a few days [41].

5 Consequently, there is a need for pharmaceutical compositions that
6 can overcome these limitations of the prior art. Such a composition
7 should maintain the stability of the active compound, preferably at both
8 room and body temperature (25 and 37°C), and exist in at least a flowable
9 state for injection, incorporation into delivery systems designed for
10 immediate, delayed, or long term administration or other means of
11 administration.

12 SUMMARY OF THE INVENTION

13
14
15 The present invention provides stable non-aqueous formulations
16 which are suspensions of peptides, polypeptides, proteins and other
17 proteinaceous substances ("proteins" or "proteinaceous substances") or
18 DNA- and RNA-containing compositions ("nucleic acids") in anhydrous,
19 aprotic, hydrophobic, non-polar vehicles with low reactivity. More
20 specifically, the present invention relates to stable formulations wherein
21 the proteinaceous substance or nucleic acid remains in stable, dry powder
22 form, yet the formulation is flowable and, therefore amenable to delivery
23 to an animal via, for example, injection, ambulatory infusion or an
24 implantable device for sustained delivery. These stable formulations may
25 be stored at elevated temperatures (e.g., 37°C) for long periods of time
26 and are especially useful as flowable formulations which can be shipped
27 and/or stored at high temperatures or in implantable delivery devices for
28 long term delivery (e.g., 1-12 months or longer) of drug.

29 In one aspect, the invention provides stable protein compositions
30 comprising a proteinaceous powder wherein the protein hydration in said
31 powder is less than about 10%; and at least one anhydrous, aprotic,
32 hydrophobic, non-polar, low-reactivity vehicle. In a preferred embodiment,

1 up to about 30% (w/w) proteinaceous powder may be used in these
2 flowable compositions.

3 In another aspect, the invention provides methods for preparing
4 stable protein compositions, said methods comprising suspending a
5 proteinaceous powder with protein hydration less than about 10% in at
6 least one anhydrous, aprotic, hydrophobic, non-polar, low-reactivity
7 vehicle.

8 In a further aspect, the invention provides methods for treating a
9 subject suffering from or susceptible to a condition which may be
10 alleviated or prevented by administration of a proteinaceous compound,
11 said methods comprising administering to said subject an effective
12 amount of a stable protein composition comprising a proteinaceous
13 powder wherein the protein hydration in said powder is less than about
14 10%; and at least one anhydrous, non-polar, aprotic, hydrophobic, low-
15 reactivity vehicle.

16 In yet a further aspect, the invention provides stable nucleic acid
17 compositions comprising a nucleic acid-containing powder wherein the
18 nucleic acid hydration in said powder is less than about 10%; and at least
19 one anhydrous, non-polar, aprotic, hydrophobic, low-reactivity vehicle.

20 In yet still another aspect, the invention provides methods for
21 preparing stable nucleic acid compositions, said methods comprising
22 suspending a nucleic acid-containing powder with nucleic acid hydration
23 less than about 10% in at least one anhydrous, non-polar, aprotic,
24 hydrophobic, low-reactivity vehicle.

25 In yet still a further aspect, the invention provides methods for
26 treating a subject suffering from or susceptible to a condition which may
27 be alleviated or prevented by administration of a nucleic acid-containing
28 compound, said methods comprising administering to said subject an
29 effective amount of a stable nucleic acid composition comprising a nucleic
30 acid-containing powder wherein the nucleic acid hydration in said powder
31 is less than about 10%; and at least one anhydrous, non-polar, aprotic,
32 hydrophobic, low-reactivity vehicle.

DETAILED DESCRIPTION OF THE INVENTION

1
2
3 The present invention is drawn to the unexpected discovery that
4 suspending dry protein- or nucleic acid-containing particles in anhydrous,
5 aprotic, hydrophobic, non-polar vehicles of low reactivity results in stable
6 flowable non-aqueous formulations. Previously known formulations of
7 proteinaceous compounds, which are dilute buffered aqueous solutions
8 containing excipients such as EDTA or polysorbate 80 which must be
9 stored at low temperatures (4-25°C), or lyophilized powders or particles
10 which must often be stored at low temperature and must then be
11 reconstituted before administration, form degradation products using
12 degradation pathways such as acid/base catalyzed hydrolysis,
13 deamidation, racemization and oxidation. Similarly, previously known
14 formulations of nucleic acids, even those prepared from lyophilized
15 powders, are administered as dilute aqueous solutions which are not
16 stable for long periods of time and which must be stored at low
17 temperatures. In contrast, the presently claimed formulations stabilize
18 proteins and nucleic acid compounds at elevated temperatures (e.g.,
19 37°C) and at high concentrations (i.e., up to about 30%).

20 Standard peptide and protein formulations consist of dilute aqueous
21 solutions. Drug stability is usually achieved by varying one or more of the
22 following: pH, buffer type, ionic strength, excipients (EDTA, polysorbate
23 80, etc). For these formulations, degradation pathways requiring water
24 (hydrolysis, deamidation, racemization) cannot be fully stabilized. In
25 contrast, in the present invention, proteinaceous compounds formulated in
26 non-aqueous, anhydrous, aprotic, hydrophobic, non-polar vehicles with
27 low-reactivity, such as mineral oil (MO), perfluorodecalin (PFD),
28 methoxyflurane (MF), perfluorotributylamine (PTA) and tetradecane (TD),
29 were shown to be chemically and physically more stable than those
30 formulated in aqueous solution. MO, PFD, MF, PTA and TD are
31 considered anhydrous, aprotic, hydrophobic, non-polar vehicles of low
32 reactivity. Such vehicles decrease the rate of degradation since they

1 isolate the proteinaceous compounds from water and they lack the ability
2 to contribute protons or other reactive moieties to degradation reactions.

3 The invention consists of using anhydrous, aprotic, non-polar,
4 hydrophobic vehicles with low reactivity such as MO, PFD, MF, PTA or
5 TD to stabilize protein formulations against both chemical and physical
6 degradation. The discovery consists of the realization that use of MO,
7 PFD, MF, PTA or TD improves the overall stability of proteins in a wide
8 range of formulation conditions, including high concentrations and
9 elevated temperatures, thus making possible shipping and/or storage of
10 protein formulations at ambient temperature and the delivery of proteins in
11 long term implantable devices that would not otherwise be feasible. The
12 present invention provides flowable pharmaceutical formulations of
13 proteinaceous substances that exhibit the requisite protein stability.
14 These non-aqueous formulations comprise two components: 1) a protein
15 in a stabilized powder formulation of low protein hydration; and 2) an
16 anhydrous, hydrophobic, aprotic, non-polar vehicle of low reactivity and
17 solubility power towards protein compounds. Optionally, the dry powder
18 form of the protein may contain stabilizers and other excipients. Such
19 stabilizers and excipients are those that further reduce protein hydration or
20 protect from interfacial tension or other dehydration process-specific
21 destabilization known to those skilled in the art.

22 Among other factors, it has been surprisingly discovered that when
23 dispersed in certain vehicles, protein powders can display significantly
24 greater stability than that observed for the dry powder alone. Such
25 vehicles include long-chain alkanes, most preferably perfluorinated forms
26 of alkanes. The present invention is especially advantageous because it
27 provides the capacity to store proteins under ambient conditions for
28 extended periods or to deliver proteins from implantable pumps for
29 extended durations.

30 Lipid/DNA and lipid/RNA complexes facilitate nucleic acid uptake
31 into cells both in vitro and in vivo. However, such complexes are
32 inherently unstable in solution, losing most, if not all, of their activity after

1 only a few days at ambient temperatures. This feature severely limits
2 their applicability for use in such devices as implantable pumps, depot
3 injection or other sustained release delivery systems where prolonged
4 residence at 37°C is needed. Lyophilization of these complexes results in
5 more stable compositions, but such powders require reconstitution prior to
6 administration to render them flowable, and the reconstituted solutions
7 are not stable. The present invention provides flowable pharmaceutical
8 formulations of nucleic acids that exhibit the requisite stability. These
9 non-aqueous formulations comprise two components: 1) a nucleic acid in
10 a stabilized powder formulation of low hydration; and 2) an anhydrous,
11 hydrophobic, aprotic, non-polar vehicle of low reactivity and solubility
12 power towards nucleic acids. Optionally, the dry powder form of the
13 nucleic acid may contain the nucleic acid in the form of lipid/DNA
14 complexes, liposomes, ribozymes, viral vectors, virosomes, dendrimers,
15 cationic polymers, PLGA particles or the like, and/or may optionally
16 contain stabilizers and other excipients. Such stabilizers and excipients
17 are those that further reduce hydration or protect from interfacial tension
18 or other process-specific destabilization known to those skilled in the art.

19 The formulations of the present invention are useful in a variety of
20 delivery systems, including, but not limited to, various pumping devices
21 (syringes, infusion sets, syringe pumps, implantable pumps, etc.),
22 transdermal reservoir systems, liquid fill capsules, injectable depot
23 compositions and the like. An additional advantage of the present
24 invention over the prior art is that the formulations of the present
25 invention prevent back diffusion of water vapor (and subsequent
26 hydrolytic degradation) because the hydrophobic vehicle of the
27 formulation acts as a barrier to water vapor. This is especially important
28 when the formulations are used in implantable devices which must remain
29 in an aqueous environment at elevated temperatures for long periods of
30 time.

31 A further advantage of the present invention is that it allows for the
32 formulation of proteins or nucleic acids in a flowable state at extremely

1 high concentrations (up to about 30% w/w). Because the protein or
2 nucleic acid is in a dry state, it is not subject to the degradation processes
3 (e.g., aggregation, precipitation or fragmentation) observed for high
4 concentration aqueous solutions.

5
6 A. Definitions:

7 As used herein, the following terms have the following meanings:

8 The term "chemical stability" means that an acceptable percentage
9 of degradation products produced by chemical pathways such as
10 oxidation, hydrolysis or enzymatic action is formed and/or that acceptable
11 biological activity is retained. In particular, a formulation is considered
12 chemically stable if no more than about 40% breakdown products are
13 formed and/or at least 40% biological activity is retained after one week
14 at 37°C.

15 The term "physical stability" means that an acceptable percentage
16 of aggregates (e.g., dimers, trimers and larger forms) and/or cleavage
17 products is formed. In particular, a formulation is considered physically
18 stable if no more than about 10% aggregates and/or cleavage products are
19 formed after one week at 37°C.

20 The term "stable formulation" means that at least about 50%
21 chemically and physically stable protein or nucleic acid compound remains
22 after one week at 37°C. Particularly preferred formulations are those
23 which retain at least about 65%, and most particularly, at least about
24 80% chemically and physically stable compound under these conditions.
25 Especially preferred stable formulations include those which remain
26 flowable at high protein or nucleic acid loading (e.g., at or above 1%).

27 The terms "protein" and/or "proteinaceous compound" and/or
28 "proteinaceous substance" mean peptides, polypeptides, proteins, viruses,
29 antibodies, etc. which comprise polymers of amino acid residues bound
30 together by amide (CONH) linkages. Both naturally-derived or purified and
31 recombinantly produced moieties are included in these terms. These
32 terms also include lipoproteins and post-translationally modified forms,

1 e.g., glycosylated proteins. Analogs, derivatives, agonists, antagonists
2 and pharmaceutically acceptable salts of any of these are included in
3 these terms. The terms also include proteins and/or protein compounds
4 and/or protein substances which have D-amino acids, modified,
5 derivatized or non-naturally occurring amino acids in the D- or L-
6 configuration and/or peptomimetic units as part of their structure.

7 The term "excipient" means a more or less inert component which
8 is added to the finished formulation other than the therapeutic ingredient.

9 The term "non-polar vehicle" means a vehicle which has a dielectric
10 constant of less than or equal to about 15.

11 The term "aprotic vehicle" means a vehicle which does not contain
12 acidic hydrogen (i.e., a hydrogen attached to an oxygen or nitrogen).

13 The term "anhydrous vehicle" means a vehicle which does not
14 contain water, including water adsorbed on its surface or combined as
15 water of crystallization.

16 The terms "vehicle with low reactivity" and/or "low-reactivity
17 vehicle" mean a vehicle which generally does not solubilize or otherwise
18 react with proteinaceous compounds and/or nucleic acids. Low-reactivity
19 vehicles are non-polar and have a Hildebrandt number of less than about
20 8.0. Examples of vehicles with low reactivity include: a) saturated
21 hydrocarbons, b) halogenated saturated or unsaturated hydrocarbons, and
22 c) esters and ethers of a) or b).

23 The terms "proteinaceous particle" and/or "proteinaceous powder"
24 mean particles which contain proteins, proteinaceous compounds or
25 proteinaceous substances. The proteinaceous particles of the present
26 invention may, optionally, contain excipients, as defined above. Such
27 excipients may include carbohydrates, non-ionic surfactants, buffers,
28 salts, carrier proteins, preservatives and the like. However, the
29 proteinaceous powders of the present invention do not contain polymers,
30 nor are they encapsulated by polymeric materials (i.e., they are not
31 microparticles or microcapsules as defined, for example, in U.S. Patent
32 No. 5,518,731).

1 The term "hydration" means water molecules associated with either
2 the protein or nucleic acid, excipients or carriers.

3 The term "hydrophobic" means incapable of dissolving to any
4 appreciable extent in water.

5 The term "nucleic acid" means unbranched (linear or circular) chains
6 of nucleotides in which the 5' phosphoric group of each nucleotide is
7 esterified with the 3' hydroxyl of the adjoining nucleotide. The term
8 includes ribonucleic acid (RNA) and deoxyribonucleic acid (DNA)
9 constructs. The term nucleic acid includes single and double stranded
10 molecules, oligonucleotides, gene expression constructs, mRNA
11 molecules, ribozymes, and the like. Naturally-derived or purified,
12 synthetically produced and recombinantly produced moieties are all
13 included in the term. The term also includes analogs, derivatives, and
14 constructs that include promoter, leader, signal, polyadenylation or intron
15 sequences, locus control regions, markers, and the like. Nucleic acids
16 containing modified, derivatized or non-naturally occurring nucleotide units
17 as part of their structure are also included in the term.

18 The terms "lipid/DNA complex" and "lipid /RNA complex" mean
19 complexes that form between nucleic acids and small, cationic unilamellar
20 vesicles held together by electrostatic interactions rather than by
21 encapsulation of the nucleic acids in liposomes. A variety of topological
22 arrangements can occur, such as DNA condensation, liposome
23 aggregation and fusion.

24 The term "liposome" means the multi- or unilamellar vesicles
25 formed from phospholipids which are used as carriers for drugs and
26 macromolecules, especially nucleic acids.

27 The terms "nucleic acid particle" and/or "nucleic acid powder"
28 mean particles which contain DNA or RNA. The nucleic acid may
29 optionally be complexed with lipids or in liposomes, ribozymes, viral
30 vectors, virosomes, dendrimers, cationic polymers, PLGA particles, or the
31 like. The nucleic acid particles of the present invention may, optionally,
32 contain excipients, as defined above. Such excipients may include

1 carbohydrates, non-ionic surfactants, buffers, salts, carrier proteins,
2 preservatives and the like.

3

4 B. Preparation of Formulations:

5 The present invention is drawn to non-aqueous formulations of
6 proteinaceous particles and nucleic acid particles with less than about
7 10% hydration suspended in anhydrous, aprotic, hydrophobic, non-polar
8 vehicles with low reactivity, which formulations are stable for prolonged
9 periods of time, even at elevated temperatures. Standard dilute aqueous
10 peptide and protein formulations require manipulation of buffer type, ionic
11 strength, pH and excipients (e.g., EDTA and ascorbic acid) to achieve
12 stability. Standard nucleic acid formulations require formulation or
13 reconstitution immediately prior to administration. In contrast, the
14 claimed formulations achieve stabilization of protein or nucleic acid
15 compounds by the use of dry particles and hydrophobic, anhydrous, non-
16 polar, aprotic low-reactivity vehicles. In particular, stability and flowability
17 of high concentrations (up to about 30%, w/w) of compound has been
18 provided by the formulations of the present invention.

19 Examples of proteins and proteinaceous compounds which may be
20 formulated using the present invention include those proteins which have
21 biological activity or which may be used to treat a disease or other
22 pathological condition. They include, but are not limited to growth
23 hormone, Factor VIII, Factor IX and other coagulation factors,
24 chymotrypsin, trypsinogen, alpha-interferon, beta-galactosidase, lactate
25 dehydrogenase, growth factors, clotting factors, enzymes, immune
26 response stimulators, cytokines, lymphokines, interferons,
27 immunoglobulins, interleukins, peptides, somatostatin, somatotropin
28 analogues, somatomedin-C, Gonadotropic releasing hormone, follicle
29 stimulating hormone, luteinizing hormone, LHRH, LHRH analogues such as
30 leuprolide, nafarelin and goserelin, LHRH agonists and antagonists, growth
31 hormone releasing factor, calcitonin, colchicine, gonadotropins such as
32 chorionic gonadotropin, oxytocin, octreotide, somatotropin plus an amino

1 acid, vasopressin, adrenocorticotrophic hormone, epidermal growth factor,
2 prolactin, somatotropin plus a protein, cosyntropin, lypressin, polypeptides
3 such as thyrotropin releasing hormone, thyroid stimulation hormone,
4 secretin, pancreozymin, enkephalin, glucagon, endocrine agents secreted
5 internally and distributed by way of the bloodstream, and the like. Further
6 agents that may be delivered include α_1 antitrypsin, insulin and other
7 peptide hormones, adrenal cortical stimulating hormone, thyroid
8 stimulating hormone, and other pituitary hormones, interferon α , β , and γ ,
9 consensus interferon, erythropoietin, growth factors such as GCSF, GM-
10 CSF, insulin-like growth factor 1, tissue plasminogen activator, CF4,
11 dDAVP, tumor necrosis factor receptor, pancreatic enzymes, lactase,
12 interleukin-1 receptor antagonist, interleukin-2, tumor suppresser proteins,
13 cytotoxic proteins, retroviruses and other viruses, viral proteins,
14 antibodies, recombinant antibodies, antibody fragments and the like.

15 Examples of nucleic acid compounds which may be formulated
16 using the present invention include those nucleic acids which code for
17 proteins which have biological activity or which may be used to treat a
18 disease or other pathological condition, such as the protein compounds
19 listed above. Nucleic acids, including sense or antisense oligonucleotides,
20 which block or reduce production of unwanted proteins are also useful in
21 the present invention. Also included in nucleic acids which may be used
22 in the present invention are those which, either directly or by coding for a
23 protein, stimulate an animal to produce immunity against a disease
24 condition (e.g., cancer) or infection by a pathogenic organism such as a
25 bacteria, virus or protozoa.

26 The above agents are useful for the treatment or prevention of a
27 variety of conditions including but not limited to hemophilia and other
28 blood disorders, growth disorders, diabetes, leukemia, hepatitis, renal
29 failure, HIV infection, hereditary diseases such as cerebrosidase deficiency
30 and adenosine deaminase deficiency, hypertension, septic shock,
31 autoimmune diseases such as multiple sclerosis, Graves disease, systemic
32 lupus erythematosus and rheumatoid arthritis, shock and wasting

1 disorders, cystic fibrosis, lactose intolerance, Crohn's disease,
2 inflammatory bowel disease, gastrointestinal and other cancers. Analogs,
3 derivatives, antagonists, agonists and pharmaceutically acceptable salts of
4 the above may also be used.

5 The protein and nucleic acid compounds useful in the formulations
6 and methods of the present invention can be used in the form of a salt,
7 preferably a pharmaceutically acceptable salt. Useful salts are known to
8 those of skill in the art and include salts with inorganic acids, organic
9 acids, inorganic bases or organic bases. Nucleic acids may also be
10 complexed with lipids or be presented as liposomes, ribozymes, viral
11 vectors, virosomes, dendrimers, cationic polymers, PLGA particles, or the
12 like.

13 The proportion of protein or nucleic acid may vary depending on the
14 compound, the condition to be treated or prevented, the expected dose
15 and the route and duration of administration. (See, for example, The
16 Pharmacological Basis of Therapeutics, Gilman et al., 7th ed. (1985) and
17 Pharmaceutical Sciences, Remington, 18th ed. (1990), the disclosures of
18 which are incorporated herein by reference.) Applicable routes include
19 oral, enteral, transdermal, percutaneous, parenteral, mucosal and the like,
20 all of which are known to those of skill in the art. The concentration of
21 protein or nucleic acid in high concentration formulations may range from
22 at least about 1% (w/w) up to about 30% while still maintaining
23 flowability. A preferred range for proteins is from about 10% to about
24 30% (w/w).

25 The vehicles useful in the present invention are non-aqueous,
26 anhydrous, aprotic, non-polar, hydrophobic vehicles with low reactivity.
27 Such vehicles have a dielectric constant less than or equal to about 15; do
28 not contain acidic hydrogen, i.e., hydrogen attached to an oxygen or
29 nitrogen; and generally do not solubilize or otherwise react with
30 proteinaceous compounds. Preferred vehicles include: a) saturated
31 hydrocarbons, b) halogenated saturated or unsaturated hydrocarbons, and
32 c) esters and ethers of a) or b). Particularly preferred vehicles are

1 perhalohydrocarbons and unsubstituted saturated hydrocarbons. Most
2 preferred vehicles are biocompatible, such as perfluorodecalin,
3 perfluorobutylamine, perfluorotripropylamine, perfluoro-N-
4 methyldecahydroquinidine, perfluoro-octahydro quinolidine, perfluoro-N-
5 cyclohexylpyridine, perfluoro-N,N-dimethylcyclohexyl methylamine,
6 perfluoro-dimethyl-adamantane, perfluorotri-methylbicyclo (3.3.1) nonane,
7 bis(perfluorohexyl) ethene, bis(perfluorobutyl) ethene, perfluoro-1-butyl-2-
8 hexyl ethene, tetradecane, methoxyflurane or mineral oil.

9 The proteinaceous or nucleic acid powders useful in the present
10 invention are solid particles wherein the hydration of the particle is less
11 than about 10% (w/w water/compound). In contrast to previous protein
12 formulations, where hydration and flexibility were required in order to
13 maintain enzymatic activity, the proteins of the particles used in the
14 present invention have minimal flexibility and minimal exposure to the
15 degradative effects of moisture since protein hydration is minimized. In
16 contrast to previous nucleic acid formulations, which required hydration in
17 order to administer the formulation, the present formulations reduce
18 hydration and degradation of the nucleic acid compounds while providing
19 a flowable formulation suitable for administration. The powders may be
20 produced by milling, spray drying, spray freeze-drying, lyophilization,
21 precipitation, and the like. These protected powder particulates are
22 preferably prepared using solid processing. They may optionally include
23 protecting agents such as carbohydrates, sucrose, trehalose, sorbitol,
24 raffinose, dextrans or cyclodextrins which may, for example, hydrogen
25 bond to the proteins to reduce their effective hydration. They may also
26 contain bulking agents such as glycine or mannitol that modify the
27 morphology and/or processing characteristics of the proteins or nucleic
28 acids, buffers that modify the pH, and non-ionic surfactants which protect
29 from surface absorption and solubilize the protein or nucleic acids. The
30 formulation of dry protein or nucleic acid powders is well known to those
31 skilled in the art.

1 Here, protein or nucleic acid hydration refers to the fraction of the
2 total moisture in a powder formulation associated with the protein or
3 nucleic acid. Certain excipients (e.g., carbohydrates) reduce the amount
4 of water associated with proteins [14] or nucleic acids. For purposes of
5 this application, protein or nucleic acid hydration will be equal to the
6 moisture content of the powder (determined, for example, by Karl Fischer
7 analysis), expressed as a percentage, multiplied by the fractional weight
8 of protein or nucleic acid in the powder.

9 Generally, the stable formulations of the present invention may be
10 prepared by simply suspending the desired amount, which may be a
11 therapeutically effective amount, of the desired proteinaceous or nucleic
12 acid powder in the selected vehicle. Preferred vehicles include MO, PFD,
13 MF, PTA and TD.

14 C. Methodology:

15 We have found that stable non-aqueous formulations of protein or
16 nucleic acid compounds may be prepared by suspending dry (less than
17 about 10% hydration) particles containing the protein or nucleic acid
18 compound to be formulated in anhydrous, aprotic, hydrophobic, low-
19 reactivity vehicle.

20 We have tested these formulations for stability by subjecting them
21 to aging at elevated temperature (37°C) and measuring the chemical
22 and/or physical stability of the formulations. Results of these studies
23 (shown, for example, in Examples 1, 2 and 3) demonstrate that these
24 formulations were stable for at least one month at 37°C.

25 A major aspect of the invention is that the flowable non-aqueous
26 formulations of the present invention are chemically and physically stable
27 at high temperatures for long periods of time. Such formulations are
28 stable even when high concentrations are used. Thus, these formulations
29 are advantageous in that they may be shipped and stored at temperatures
30 at or above room temperature for long periods of time. They are also
31 suitable for use in implantable delivery devices.

1 **DISCLOSURE OF EXAMPLES OF THE INVENTION**

2
3 The following method was used to perform the studies in the
4 Examples that follow.

5 **Karl Fischer Moisture Analysis :** Vials and stoppers were dried
6 overnight in a vacuum oven at 80°C. Approximately 6 mg of sample was
7 weighed into a dry vial and the vial was stoppered. Control vials were
8 prepared by simply stoppering an empty dry vial (i.e., a vial containing no
9 sample). Subsequently, 150 μ L aliquots of dry methanol was added to
10 sample and control vials via a 250 μ L Hamilton Syringe (Hamilton Co.,
11 Reno, NV) which had been previously washed three times with dry
12 methanol. The vials were then sonicated at room temperature until all
13 solids were dispersed, centrifuged, and 100 μ L of the supernatant
14 methanol was injected into an Aquatest 10 Coulometric Moisture Analyzer
15 (SeraDyn Inc., Indianapolis, IN). The resultant readings were recorded,
16 and water content of the sample calculated by subtracting the control
17 reading from that of the sample.

18 The following reagents were used to perform the studies in the
19 Examples that follow.

20 Perfluorodecalin, perfluorotributylamine and tetradecane were
21 purchased from Aldrich Chemical Company (Milwaukee, WI).
22 Methoxyflurane was purchased from Abbott Laboratories (North Chicago,
23 IL). Light Mineral Oil USP was purchased from Spectrum Chemical Corp.
24 (Gardena, CA).

25 The following examples are offered to illustrate this invention and
26 are not meant to be construed in any way as limiting the scope of this
27 invention.

EXAMPLE 1**ALPHA-INTERFERON FORMULATIONS****STABILITY OF ALPHA INTERFERON (α -IFN) SUSPENSIONS**

Human recombinant Interferon- α -2a (α -IFN) (Scitech Genetics Ltd., lot# 036R2801) was formulated as a 5 mg/mL solution containing 5mM citrate, 0.5% sucrose, and 0.005% Tween 80, pH 4.5. Aliquots of 200 μ L of this solution were then dispensed into 1 mL glass lyophilization vials, partially covered with lyophilization stoppers, and lyophilized using an FTS Systems lyophilizer according to the following cycle:

Pre-cool shelves to 5°C;

Load vials;

Freeze to -50°C at 2.5°C/min;

When product is at -30°C set vacuum to 125 mT;

Hold at -50°C for 30 min;

Ramp to 0°C at 0.5°C/min;

Hold at 0°C for 120 min;

Ramp to 20°C at 1°C/min;

Hold at 20°C for 120 min;

Ramp to 30°C at 1°C/min;

Hold at 20°C for 1000 min; and

Stopper vials.

The resultant powder had a moisture content of approximately 5% (w/w) as determined by Karl Fischer analysis and a protein hydration of about 2.5%. Suspensions were prepared by adding 100 μ L of either perfluorodecalin (PFD), methoxyflurane (MF), or mineral oil (MO) to the vials containing the α -IFN powder, and the vials incubated at 37°C. Samples were pulled at 2 and 4 weeks, and the α -IFN extracted from the non-aqueous phase by adding 700 μ L of buffer (containing 5mM citrate, 0.5% sucrose, and 0.005% Tween 80, pH 4.5) and gently inverting the vials. After 15 minutes, an aliquot of the aqueous phase was removed and analyzed for stability by reverse phase HPLC and reduced and non-

1 reduced SDS-PAGE electrophoresis.

2 The formulations remained chemically stable as determined by
3 reverse phase HPLC (Table 1). In addition, no aggregation or cleavage
4 products were observed on reduced or non-reduced SDS-PAGE gels.

5 **Table 1**

6 **Stability of α -IFN suspensions at 37°C as measured by reverse phase chromatography**

Time (weeks)	% Recovery PFD susp 37°C	% Recovery MF susp 37°C	% Recovery MO susp 37°C
0	98 \pm 3	92 \pm 6	101 \pm 1
2	103 \pm 2	81 \pm 3	94 \pm 3
4	98 \pm 1	81 \pm 1	84 \pm 2

7 * Numbers represent mean \pm standard deviation of 2-3 samples

8

9 **RP-HPLC**

10 Instrument: Hewlett Packard HP-1090
11 Flow Rate: 0.3 mL/min
12 Detection: 210 nm
13 Column: Waters Delta-Pak, c18, 150 x 2 mm, 300Å.
14 Mobile Phase: A = 30/70/0.2 Acetonitrile/Water/TFA
15 B = 80/20/0.2 Acetonitrile/Water/TFA

16

17 Gradient:

<u>Time</u>	<u>%B</u>
18 0	23
19 45	35
20 55	52
21 60	90
22 65	90
23 68	23

24

25 **SDS-PAGE**

26 Apparatus: Life Technologies Vertical Gel Electrophoresis
27 system.
28 Gel: 15% discontinuous, 15 x 17 cm, 0.8 mm thick.
29 Running Conditions: 200v, 50 mA, approximately 3 hrs.
30 Staining: Coomassie Blue R-250
31 Gel Analysis: Bio-Rad GS-700 image analyzer with Molecular
32 Analyst software.
33

EXAMPLE 2**STABILITY OF CHYMOTRYPSIN FORMULATIONS**

Formulations were prepared containing 2% chymotrypsin (Worthington Biochemical Corp., 1X Crystallized, Lot# H5B7405L), determined by Karl Fischer analysis to have a water content and protein hydration of approximately 7% (w/w), either dissolved in 0.1M borate buffer, pH 8.0, or suspended (as a dry powder) in either perfluorodecalin or Light Mineral Oil, U.S.P. Samples were stored at 37°C for 10 weeks, and assayed for chymotrypsin activity using casein as a substrate.

The results are shown in Table 2 and demonstrate stability of the formulations.

Chymotrypsin Bioactivity assay

Samples were diluted in 0.1 M borate buffer, pH 8.0, such that the final chymotrypsin concentration for assay was approximately 2 - 50 $\mu\text{g/mL}$. A casein substrate solution was prepared by suspending 1 gm of casein in 95 mL borate buffer, pH 8.0 and heating in a boiling water bath until the casein had dissolved (approximately 10 minutes), then adding 1.1 mL 5% CaCl_2 and diluting the solution to 100 mL with 0.1M borate buffer solution at pH 8.0. The substrate solution (1.0 mL) was prewarmed at 37°C in a heating block, and to it was added 1.0 mL of the sample. The solutions were mixed and incubated at 37°C for exactly 20 minutes. Subsequently, 3.0 mL of 5% trichloroacetic acid was added, and the resultant mixture was allowed to stand at room temperature for 30 minutes, then centrifuged for 20 minutes at 3,000 g. The absorbance of the supernatant was read in a UV spectrophotometer at 280 nm and the activity (in Units/mg) calculated by the following equation.

$$\text{Activity} = \frac{A_i}{(C)(t)}$$

1 where: A_t = absorbance of supernatant (at 280 nm) at time t of the
2 reaction (in this case, 20 minutes); C = concentration of chymotrypsin in
3 sample; and t = time of reaction (20 minutes).

4 **Table 2**

5 **Activity of chymotrypsin formulations when stored at 37°C**

Time (weeks)	% Remaining PFD*	% Remaining MO*	% Remaining Buffer*
0	103 ± 5	100 ± 5	100 ± 4
1	97 ± 2	86 ± 1	23 ± 2
3	102 ± 3	96 ± 3	19 ± 2
6	102 ± 2	89 ± 1	22 ± 4
10	102 ± 3	92 ± 2	

6 * Numbers represent mean ± standard deviation of 6 samples

7 **EXAMPLE 3**

8 **STABILITY OF PLASMA PROTEIN SUSPENSIONS**

9
10 Formulations of a post-translationally modified plasma protein of 55
11 kilodalton molecular weight were prepared containing 1 mg/mL protein
12 and approximately 30 mg/mL excipients, buffered to a neutral pH. One
13 mL aliquots of the above solutions were pipetted into 3 mL glass vials,
14 covered with lyophilization stoppers, loaded into a freeze dry chamber
15 (FTS Systems Inc.), and lyophilized.

16 The resultant powder had a final moisture content of about 0.25%
17 (w/w) water, as determined by Karl Fischer analysis and protein hydration
18 of about 0.008%. Suspensions were prepared by adding 1 mL of either
19 perfluorodecalin (PFD) or methoxyflurane (MF) to the vials containing the
20 dry protein powder, and the vials were then incubated at 37°C. Control
21 samples of the lyophilized powder were stored at -80°C. Samples were
22 pulled at 0, 4.5, 6.5, 8.5 and 12.5 weeks and analyzed for activity using
23 a bioactivity assay, and for chemical stability by size exclusion
24 chromatography.

25 The results are summarized in Tables 3 and 4 and show that the
26 formulations remained chemically (as determined by biological activity)

1 and physically (as determined by SEC) stable.

2 **Size Exclusion Chromatography**

3 Column: TSD G3000 swxl column, 7.8 x 300 mm, 5 μ m
 4 (ToSoHaas TO8541 or equivalent)
 5 Mobile Phase: 50 mM Na₂HPO₄, 150 mM NaCl, pH 7.0
 6 Flow Rate: 1.0 mL/min
 7 Detector: 214 nm
 8 Injection Volume: 50 μ L

9
10 **Table 3**

11 **Stability of plasma protein suspensions at 37°C as measured by bioactivity assay**

Time	% LS†	%LS†	%LS†
(weeks)	Lyo. Powder	PFD susp. 37°C	MF susp
	-80°C		37°C
0	92 \pm 14	84 \pm 14	92 \pm 4
1.5	98 \pm 12	109 \pm 9	107 \pm 16
4.5	89 \pm 2	86 \pm 4	61 \pm 20
6.5	94 \pm 7	101 \pm 0	68 \pm 15
8.5	110 \pm 2	97 \pm 2	62 \pm 5
12.5	111 \pm 7	105 \pm 11	

12 †%LS = % Label Strength = $\frac{[\text{protein}] \text{ test}}{[\text{protein}] \text{ control}}$

13
14 * Numbers represent mean \pm standard deviation of 3 samples

15
16 **Table 4**

17 **Stability of plasma protein suspensions at 37°C as measured by size exclusion**
 18 **chromatography.**

19

Time	% LS†	%LS†	%LS†
(weeks)	Lyo. Powder	PFD susp.	MF susp
	-80°C*	37°C*	37°C*
0	92 \pm 0	96 \pm 1	84 \pm 7
1.5	107 \pm 3	106 \pm 2	104 \pm 4
4.5	108 \pm 2	96 \pm 1	67 \pm 35
6.5	113 \pm 2	101 \pm 2	79 \pm 12

8.5	105 ± 1	95 ± 4	57 ± 5
12.5	100 ± 3	98 ± 1	

1 †%LS = % Label Strength = $\frac{[\text{protein}] \text{ test}}{[\text{protein}] \text{ control}}$

2

3 * Numbers represent mean ± standard deviation of 3 samples

4 EXAMPLE 4

5 HIGH CONCENTRATION FLOWABLE FORMULATIONS

6

7 Solutions were prepared containing either Albumin (Sigma, Lot
8 129FO1431), Lysozyme (Sigma Lot 65H7025) or Trypsinogen
9 (Worthington Lot# 38E273N) and sucrose in a 1:1 (w/w) ratio. The
10 solutions were spray dried on a Yamato ADL 31 Spray Dryer (Yamato
11 Corp., NY) with the following parameters: inlet temp 120°C, outlet
12 temperature 65°C, atomizer 1.2 kg/cm². The powders were then
13 transferred to a vacuum oven and allowed to further dry at 30°C overnight
14 under full vacuum. The moisture content of the powders studied was
15 approximately 4.5 % (w/w) as determined by Karl Fischer analysis with a
16 protein hydration of about 2.25%.

17 Pastes were formulated by mixing 700 mg of each powder with 1.0
18 mL of perfluorodecalin (approximately 28% w/w). The pastes were
19 loaded into 1.0 cc syringes fitted with 30 Gauge needles (Becton
20 Dickinson), and extruded. All pastes extruded evenly and completely with
21 little effort.

22 EXAMPLE 5

23 STABILITY OF FACTOR IX SUSPENSIONS

24

25 Coagulation Factor IX (FIX) from human serum (Calbiochem-
26 Novobiochem, La Jolla, CA) was formulated as a 0.5 mg/mL solution
27 containing 60 mg/mL sucrose, 60 mg/mL mannitol, 1 mg/mL polysorbate
28 80 and 1.6 mg/mL histidine buffer buffered to a pH of approximately 7.
29 One mL aliquots of this solution were lyophilized according to the cycle
30 above. The resultant powder had a moisture content of 1%, as

1 determined by Karl Fischer analysis.

2 Suspensions were prepared by adding 1 mL of perfluorodecalin
3 (PFD), perfluorotributylamine (PTA) or tetradecane (TD) to the vials
4 containing the dry FIX powder. The vials were incubated at 37°C.
5 Control samples of the lyophilized powder were stored at -80°C. Samples
6 were pulled at 0 and 2 weeks and analyzed for FIX activity by clotting
7 bioactivity assay, and for chemical stability by size exclusion
8 chromatography.

9 The results (Tables 5 and 6) showed that the formulations remained
10 chemically (as determined by biological activity) and physically (as
11 determined by SEC) stable.

12 **Table 5**
13 **Stability of Factor IX Suspensions at 37°C as Measured by**
14 **Bioactivity Assay**
15

Time	% LS†	%LS†	%LS†
(weeks)	PFD susp.	PTA susp.	TD susp.
	37°C	37°C	37°C
0	97 ± 2	89 ± 3	95 ± 3
2	98 ± 2	96 ± 1	96 ± 1

16 †%LS = % Label Strength = $\frac{[\text{protein}] \text{ test}}{[\text{protein}] \text{ control}}$
17

18 * Numbers represent mean ± standard deviation of 3 samples
19

20 **Table 6**
21 **Stability of Factor IX Suspensions at 37°C as Measured by Size**
22 **Exclusion Chromatography.**
23

Time	% LS†	%LS†	%LS†
(weeks)	PFD susp.	PTA susp.	TD susp.
	37°C	37°C	37°C
0	94 ± 1	93 ± 1	97 ± 1
2	94 ± 2	95 ± 2	96 ± 1

$$\dagger \%LS = \% \text{ Label Strength} = \frac{[\text{protein}] \text{ test}}{[\text{protein}] \text{ control}}$$

* Numbers represent mean \pm standard deviation of 3 samples

EXAMPLE 6

STABILITY OF NUCLEIC ACID SUSPENSIONS

Plasmid pCIN.CAT was made by cloning the coding sequence for bacterial chloramphenicol acetyltransferase (CAT) into the expression plasmid pCIneo (Promega). The CAT coding region was isolated by PCR amplification from plasmid pSIS.CAT [42] by standard techniques (PCR Technology, 1989. H.A. Erlich, ed. Stockton Press, incorporated herein by reference). These primers produced a unique XhoI restriction site at the 5'-end and a unique NotI restriction site at the 3' end. This fragment was subcloned into the XhoI and NotI sites of pCIneo by standard techniques (Molecular Cloning, second edition, 1989. Sambrook, J., Fritsch, E.F., and Maniatis, T., incorporated herein by reference.) Plasmid DNA was grown in bacterial culture and isolated (Qiagen, GmbH).

Formulations were prepared containing 100 mg/ml sucrose, 100 mg/ml mannitol, 10 μ g/ml pCIN-CAT DNA, 50 μ g/ml of a 1:1 formulation of DOTMA (n-[1-(2,3 dioleoyloxy)propyl]-n,n,n-trimethylammoniumchloride) and DOPE (dioleoyl phosphatidylethanolamine) (Lipofection, GIBCO BRL) in 10 mM Tris buffer at pH 7.1. Aliquots of 200 μ l of the above formulation were pipetted into 1 ml glass vials and lyophilized using the following protocol:

Precool shelf temperature to 5°C;

Load vials;

Freeze to -40°C at 0.4°C/min and hold at -40°C for 120 minutes;

Ramp to -10°C at 0.4°C/min and hold for 240 minutes;

Ramp to -45°C at 0.4°C/min and hold for 120 minutes;

Set vacuum to 100mT;

Hold at -45°C for 360 minutes with vacuum at 100 mT;

Ramp to -25°C at 0.04°C/min with vacuum at 100 mT;

1 Hold at 25°C for 1500 minutes with vacuum at 100 mT.

2 The subsequent dry powder had a moisture content of
3 approximately 2% as measured by Karl Fischer analysis. Suspensions
4 were prepared by adding 300 μ l of perfluorodecalin (PFD) to the vials in a
5 glove box under dry nitrogen. Suspension, dry powder and solution
6 samples were incubated at 37°C for 1, 4 and 7 days, and subsequently
7 monitored for biological activity by monitoring gene transfer efficiency as
8 measured by CAT expression in HEK293 cells. Transfection of HEK 293
9 cells with lipid/DNA complexes was performed as described by the
10 Manufacturer (GIBCO BRL).

11 The results are shown in Table 7, and demonstrate that when
12 lipid/DNA complexes were formulated in aqueous solution, essentially all
13 activity was lost when the solution was stored at 37°C for 1 week. In
14 contrast, both the lyophilized dry nucleic acid powder and the nucleic acid
15 powder suspended in PFD retained essentially all their biological activity
16 (within the experimental variability of the assay) when stored for 1 week
17 at 37°C.

18 **Table 7**

19 **Transfection Activity of Lipid/DNA Constructs After**
20 **Incubation at 37°C. (Numbers are mean \pm standard**
21 **deviation of 12 replicates.)**

22 **Average ng CAT Protein Expressed Per mg Total Cellular Protein**

Time (days)	Solution Formulation	Dry Powder Formulation	PFD
			Suspension Formulation
0	478 \pm 254	219 \pm 114	n.d.
1	115 \pm 46	628 \pm 192	273 \pm 122
4	13 \pm 12	255 \pm 137	284 \pm 267
7	6 \pm 3	377 \pm 202	339 \pm 151

1 Modification of the above-described modes of carrying out various
2 embodiments of this invention will be apparent to those of skill in the art
3 following the teachings of this invention as set forth herein. The
4 examples described above are not limiting, but are merely exemplary of
5 this invention, the scope of which is defined by the following claims.

1 What is claimed is:

2

3 1. A stable non-aqueous composition of an active agent comprising:

4 a) an active agent containing powder wherein the active agent

5 hydration in said powder is less than about 10%; and

6 b) at least one anhydrous, aprotic, hydrophobic, non-polar, low-

7 reactivity vehicle, wherein said active agent is selected from the group

8 consisting of proteins, proteinaceous compounds and nucleic acids.

9

10 2. The composition of Claim 1 wherein at least about 80% of the

11 active agent remains stable for at least one month at 37°C.

12

13 3. The composition of Claim 1 wherein said active agent hydration

14 is less than about 5%.

15

16 4. The composition of Claim 1 wherein said vehicle is selected from

17 the group consisting of perhalohydrocarbons, unsubstituted saturated

18 hydrocarbons, halogenated hydrocarbons, esters of unsubstituted

19 saturated or halogenated hydrocarbons and ethers of unsubstituted

20 saturated or halogenated hydrocarbons.

21

22 5. The composition of Claim 1 wherein said vehicle is selected from

23 the group consisting of MO, PFD, MF, PTA and TD.

24

25 6. The composition of Claim 1 wherein said powder comprises up

26 to about 30% (w/w) of said composition.

27

28 7. The composition of Claim 1 wherein said active agent is a

29 protein selected from the group consisting of Factor IX, Factor VIII, alpha-

30 interferon, consensus interferon, beta-galactosidase, lactate

1 dehydrogenase, chymotrypsin, trypsinogen, an antibody, and analogs
2 thereof.

3

4 8. The composition of Claim 1 wherein said active agent is a
5 nucleic acid selected from the group consisting of DNA, RNA and
6 oligonucleotides.

7

8 9. The composition of Claim 8 wherein said nucleic acid is in the
9 form of at least one selected from the group consisting of a nucleic
10 acid/lipid complex, a nucleic acid-containing liposome, a ribozyme, a viral
11 vector, a virosome, nucleic acid-containing dendrimers, nucleic acid-
12 containing cationic polymers and nucleic-acid-containing PLGA particles.

13

14 10. The composition of Claim 1 wherein said active agent is
15 pharmaceutically useful.

16

17 11. A method for preparing the composition of Claim 1 comprising
18 suspending an active agent-containing powder with active agent hydration
19 less than about 10% in at least one anhydrous, aprotic, hydrophobic, non-
20 polar, low-reactivity vehicle.

21

22 12. A method for treating a subject suffering from or susceptible to a
23 condition which may be alleviated or prevented by administration of an
24 active agent according to Claim 1, said method comprising administering
25 to said subject an effective amount of the composition of Claim 1.

26

27 13. The method of Claim 12 wherein said condition is hemophilia
28 and the active agent in said composition is selected from the group
29 consisting of Factor VIII, Factor IX, and analogs thereof.

1 14. The invention of any of Claims 1 and 11 wherein said powder
2 comprises about 10 to about 30% (w/w) of said composition.

3

4 15. The invention of any of Claims 1 and 12 wherein administration
5 of the composition is via a route selected from the group consisting of
6 parenteral, transdermal, mucosal, oral and enteral.

7

8 16. The invention of any of Claims 1 and 12 wherein administration
9 of the composition is via an implantable delivery device.

10

11 17. The invention of any of Claims 1 and 12 wherein administration
12 of the composition is long-term continuous administration.

INTERNATIONAL SEARCH REPORT

International Application No

PCT/US 97/18575

A. CLASSIFICATION OF SUBJECT MATTER

IPC 6 A61K47/06 A61K9/10 A61K38/48 A61K38/17 A61K48/00

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 6 A61K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	FR 2 342 073 A (MERIEUX INST) 23 September 1977 see page 1, line 32 - page 2, line 8 see page 5, line 16-21 see table	1-4, 10-12,15
X	EP 0 177 342 A (GENENTECH INC) 9 April 1986 see abstract see page 3, line 5-15 see page 5, line 11-25 see page 7, line 27 - page 8, line 2 see page 9, line 15 see claims 1,3-6,12-14	1,4-7, 10,12,15

☒ Further documents are listed in the continuation of box C.

☒ Patent family members are listed in annex.

* Special categories of cited documents :

A document defining the general state of the art which is not considered to be of particular relevance

E earlier document but published on or after the international filing date

L document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)

O document referring to an oral disclosure, use, exhibition or other means

P document published prior to the international filing date but later than the priority date claimed

T later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

X document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

Y document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art

a document member of the same patent family

Date of the actual completion of the international search

10 February 1998

Date of mailing of the international search report

18.02.98

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INTERNATIONAL SEARCH REPORT

International Application No
PCT/US 97/18575

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	<p>EP 0 427 998 A (PHIDEA SRL) 22 May 1991</p> <p>see abstract see page 2, line 51-54 see example 1 see table 2</p> <p>-----</p>	<p>1,11,12, 15</p>

INTERNATIONAL SEARCH REPORT

international application No.
PCT/US 97/18575

Box I Observations where certain claim were found unsearchable (Continuation of item 1 of first sheet)

This International Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☒ Claims Nos.:
because they relate to subject matter not required to be searched by this Authority, namely:
see FURTHER INFORMATION sheet PCT/ISA/210
2. ☐ Claims Nos.:
because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful International Search can be carried out, specifically:
3. ☐ Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

1. ☐ As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims.
2. ☐ As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. ☐ As only some of the required additional search fees were timely paid by the applicant, this International Search Report covers only those claims for which fees were paid, specifically claims Nos.:
4. ☐ No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest

- ☐ The additional search fees were accompanied by the applicant's protest.
- ☐ No protest accompanied the payment of additional search fees.

INTERNATIONAL SEARCH REPORT

International Application No. PCT/US 97/18575

FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

Remark : Although claims 12 and 13 are directed to a method of treatment of the human body , the search has been carried out and based on the alleged effects of the composition.

INTERNATIONAL SEARCH REPORT

Information on patent family members

International Application No

PCT/US 97/18575

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
FR 2342073 A	23-09-77	NONE	
EP 0177342 A	09-04-86	JP 61093129 A	12-05-86
EP 0427998 A	22-05-91	EP 0432431 A	19-06-91
		ES 2067622 T	01-04-95
		ES 2067624 T	01-04-95
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